

TITLE

From Page No. 80

RD of .SKO CH₂CH₃ / FUS 29 #1

#1 mixed

10 μ l MP DNA
2 μ l 10x B
0.5 μ l EcoRI
1 μ l HindIII
6.5 μ l H₂O

Inc 37°C ~ 2.5 hrs.

Stored -20°C O/N.

To Page No. 82

Witnessed & Understood by me,

Date

Invented by

Date

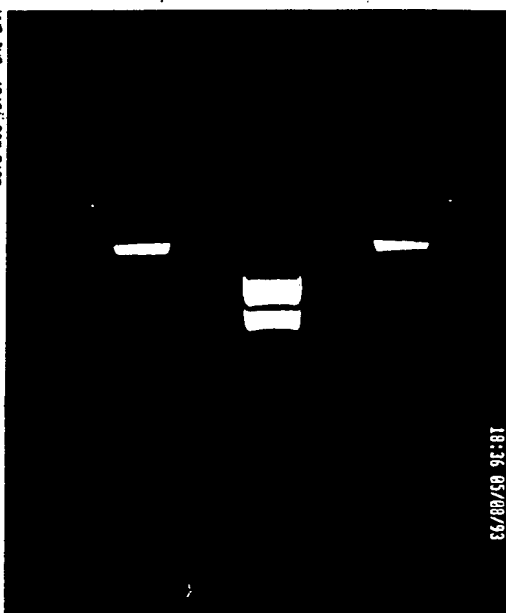
Recorded by

WTH Bauman

From Page No. 81

Runned a 1% LMP agarose gel (1x TBE)
 Ran SK0/CH2CH3 F43 29 #1 out on gel

UP Inc. (818) 285-1123



Gel is severely
 overloaded.

Indicated band should
 be running @ 2 Kb.

Cut out indicated band

Did Magic PCR prep.

Rs'd in 150 µl TE.

Ligated (Hi + Lo conc) to

pSV15.TD.LL = EcoRI/HindIII

(did vect cont too).

Inc all 3 @ 12.5°C O/N.

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Witnessed & Understood by me,

Date

Invented by

Recorded by

W. A. Baron

Date

TITLE

From Page No. 82

Transformed o/n pSV15/FUS 29 ligations
into competent E. coli
Plated each on 5x100mm LB canb 5° plates.
(control plated onto 1 plate).
Inc all (11) 37°C o/n.

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Witnessed & Understood by me,

Date

Invented by

Recorded by

Date

83

Project No. 713
Book No. 17522 TITLE _____

Exhibit E, pg. 4 of 7

84

From Page No. 83

Checked transformation plates (pSV15/Fus29)
Started 40 x 5ml LB can⁵⁰ MP's + Master
Inc all @ 37°C O/N.

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Witnessed & Understood by me,

Date

Invented by

Recorded by

Date

TITLE

From Page No. 84

Ran 40 O/N pSV15/FUS 29 MP's in Autogen

Recovered each in ~200 μ l TE

RD's

Per rxn

4 μ l MP DNA

2 μ l 10x B

0.5 μ l EcoRI

0.5 μ l HindIII

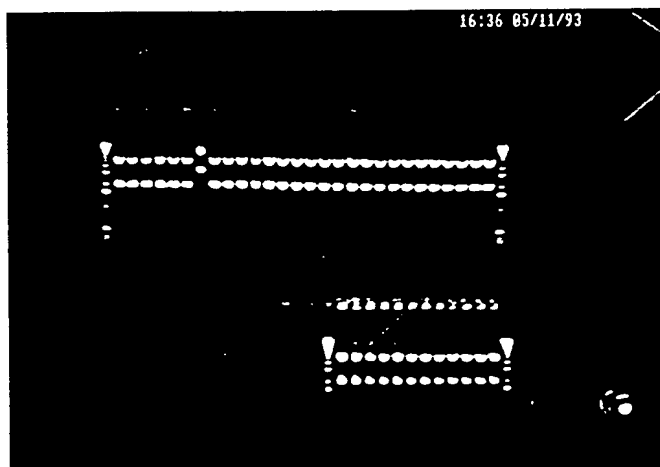
1 μ l RNaseA

12 μ l H₂O

20

Inc all (40) 37°C - 2 hrs (+ 4 μ l dye)

Run 15 μ l each on 0.7% agarose (1x TBE)



UVP INC. (818) 285-3123

÷ EcoRI

Ready to
transfect after
I do a large
scale prep.

#1
Grew in 2x 500ml 2YT
carb 5% O/N 37°C

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To Page No. 81

Witnessed & Understood by me,

Date

Invented by

Recorded by

Date

From Page No. 85

Did Magic Maxi prep on PSV15/FUS 29
EtOH ppt'd after elution from resin
Stored as ppt in 70% EtOH -20°C o/w.

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Witnessed & Understood by me,

Date

Invented by

Date

Recorded by

Will Bacon

TITLE

From Page No. 86

Spun down pSV15/FUS 29 + pSV15/HPTK6 (Full length)

Washed 3x 1ml -20°C 70% ETOH

Dried
RS'd each in 400µl TE

measured O.D₂₈₀ of 10µl each (in 1ml)

HPTK6
(Full length) = $0.382 \times 50 = 19.1 \frac{\mu\text{g}}{\text{ml}} \div 10 = 1.9$

Fus 29 = $0.416 \times 50 = 20.8 \div 10 = 2.08 \frac{\mu\text{g}}{\text{ml}}$

Picked up DP12 (CHO) cells for transfection

on [redacted]

Have 50ml
put into a spinner
flask

Added 100ml
F12 + 5% dFBS
+ 10mM HEPES
+ 1x GHT

Inc 37°C O/N.

INTERNAL TRANSMITTAL FORM			
Genentech, Inc. Genentech, Inc. Genentech, Inc. Genentech, Inc. Genentech, Inc.	Will Bacon	Will Bacon	409
C. No. 1	C. No. 2	CBC 613	
DP-12	208-215	50ml	
FORMER CONDITIONS		10594	
FROM INSTRUCTIONS			
2371p23			
BSA + 5% WBS + 1x GHT			
count 22.0x10 ⁵ C/ml (98%)			

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No. 87

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Witnessed & Understood by me,	Date	Invented by	Date
		Will Bacon	
		Recorded by	